

# TGuide Smart Hi-Plant DNA Kit

(Prefilled 96-Deepwell plate)

*For extraction of genomic DNA from plant tissues, especially plant materials rich in polysaccharides, polyphenols or starch.*

## TECHNICAL MANUAL

Cat. no. GDP617-E

**Note:** To use the TGuide Smart Hi-Plant DNA Kit, you must have the TGuide Smart Hi-Plant DNA Kit (program no. DP617) installed on the TGuide S16/S32 Pro/S96 Dex Nucleic Acid Extractor.



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For Research Use Only. Not for use in diagnostic procedures.

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# TGuide Smart Hi-Plant DNA Kit

Cat. no. GDP617-E

## Kit Contents

| Contents              | GDP617-E<br>(96 preps) |
|-----------------------|------------------------|
| Buffer GPH            | 85 ml                  |
| NKY tablets           | 5 tablets              |
| Proteinase K          | 2×1 ml                 |
| Buffer ST             | 5×1 ml                 |
| RNaseA (100 mg/ml)    | 500 µl                 |
| Buffer TB             | 15 ml                  |
| Hi-Plant DNA Reagents | 6 plates               |
| TGuide Smart Tip Comb | 12 pcs                 |

## Hi-Plant DNA Reagents composition

| Column<br>1/7 | Column<br>2/8 | Column<br>3/9 | Column<br>4/10 | Column<br>5/11 | Column<br>6/12                 |
|---------------|---------------|---------------|----------------|----------------|--------------------------------|
| Buffer DBP5   | Buffer LC4E   | Buffer BP4    | Buffer RWS6P   | None           | MagAttract<br>Suspension BESP2 |
| 240 µl        | 900 µl        | 900 µl        | 900 µl         |                | 910 µl                         |

## Storage

Store at room temperature (15-30°C) in a dry environment. If precipitates form in any buffer, incubate the solution in a 37°C water bath for 10 minutes to fully dissolve precipitates.

## Introduction

This kit adopts specially functionalized magnetic beads paired with delicately formulated buffer systems to isolate and purify high-quality genomic DNA from a wide range of plant tissues, especially plant materials rich in polysaccharides, polyphenols or starch, as well as macrofungal specimens. Optimized polyphenol removal tablets efficiently eliminate complex secondary metabolites contained in plant samples.

Compatible with fully automated nucleic acid extractors including TGuide S16 / S32 Pro / S96 Dex, the uniquely coated magnetic beads exhibit strong affinity for nucleic acids under defined buffer conditions; upon condition modulation, captured nucleic acids are eluted from beads to realize rapid nucleic acid separation and purification. Custom magnetic combs capture, transfer and release magnetic beads to complete nucleic acid translocation, enabling high-throughput automated extraction.

The whole extraction workflow is safe and user-friendly. The purified genomic DNA is characterized by high purity, large fragment size, and consistent quality. DNA obtained using this kit is suitable for a variety of downstream applications, including restriction enzyme digestion, PCR amplification, library preparation, Southern blot analysis, microarray assays, high-throughput sequencing, and third-generation sequencing.

## Features

**Broad Compatibility:** Suitable for genomic DNA extraction from a wide range of polyphenol- and polysaccharide-rich plant tissues and fungal samples.

**Safe and Non-toxic:** No toxic organic reagents such as phenol or chloroform required.

**Simple and Rapid:** Ultra-pure genomic DNA can be obtained within 1 hour.

**High Purity:** Purified DNA can be directly used for microarray analysis and high-throughput sequencing.

## Notes Please read these notes carefully before using this kit.

1. Add NKY tablets into Buffer GPH before use: add one NKY tablet into 21 ml Buffer GPH for extraction of 24 samples. Select one of the following NKY tablets application protocols as appropriate for your experiment:
  - 1) For small batch of samples: Crush the tablet into pieces (mung bean-sized pieces); add approximately 40 mg of NYK tablet piece per sample, followed by Buffer GPH and vortex for 2 min before subsequent extraction.
  - 2) For large batch of samples: Dissolve one tablet in 21 ml Buffer GPH and vortex for 2 min to form a white opaque suspension (stable for 4 weeks at ambient temperature). Mix the solution thoroughly before each use. Minor unevenness

after dispensing into centrifuge tubes does not compromise experimental performance.

2. Avoid repeated freeze-thaw cycles of samples, which will result in shortened DNA fragments and reduced DNA yield.

## Recommended Input Amounts for Various Sample Types

| Sample Type                              | Recommended Input Amount | Example Sample Library                                                                                                                                                                            |
|------------------------------------------|--------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Common Leaves                            | 60-100 mg                | Rice, Maize, Wheat, Soybean, Phoenix tree, Reed, Sweet potato                                                                                                                                     |
| Polysaccharide- & Polyphenol-Rich Leaves | 30-60 mg                 | Strawberry, Willow, Eucalyptus, Smoketree, Acer truncatum, Pomelo, Maple, Chinese rose, Chrysanthemum, Hosta, Lonicera maackii, Mulberry leaf, Hawthorn, White poplar, Toona, Bodhi tree, Weigela |
| High-Moisture Tissues                    | 100-300 mg               | Chinese cabbage, Wild peach, Muskmelon                                                                                                                                                            |
| Flower Petals                            | 60-150 mg                | Rose, Chrysanthemum, Wild rose, Yellow iris, Yellow iris pollen, Pine pollen                                                                                                                      |
| Roots & Stems                            | 100-200 mg               | Salvia miltiorrhiza root, Curcuma root, Ginseng, Maize stalk, Reed stalk                                                                                                                          |
| Tubers                                   | 50-100 mg                | Sweet potato, American ginseng                                                                                                                                                                    |
| Seeds                                    | 30-60 mg                 | Peanut, Bean sprout, Ginseng, Barley, Maize                                                                                                                                                       |
| Traditional Chinese Medicinal Materials  | 30-60 mg                 | Curcuma, Salvia miltiorrhiza, Polygonatum, Fritillary bulb                                                                                                                                        |
| Fungal Samples                           | 30-60 mg                 | Hypsizygus marmoreus, White beech mushroom, Shiitake, Enoki mushroom, Dictyophora, Aspergillus niger                                                                                              |

## Protocol

### 1. Preparation of Hi-Plant DNA Reagents

- 1.1 Remove the pre-filled 96-well deep plate from the kit. Invert it several times to resuspend the magnetic beads. Remove the packaging bag, and gently tap or shake the plate to collect the reagents and magnetic beads at the bottom of the wells (alternatively, centrifuge the plate at 500 rpm for 1 min using a plate centrifuge). Carefully peel off the sealing film before use, avoiding vibration of the plate to prevent liquid splashing.
- 1.2 Add appropriate volume (60-100  $\mu$ l) of elution Buffer TB to wells 5/11 before start.

### 2. Sample Preparation

#### 2.1 Sample Grinding

- a. Liquid nitrogen grinding: According to the recommended input amount for different sample types, take the plant tissue and grind it thoroughly in liquid nitrogen.
- b. Homogenizer processing: Snap-freeze samples in liquid nitrogen, then transfer it to the steel bead grinding tubes (TIANGEN, Cat. no. OSE-TH-B04). Homogenize with the TGrinder H24 Tissue Homogenizer (TIANGEN, Cat. no. OSE-TH-01): oscillate at a speed of 6.5 m/s for 60 sec, with a 15-second interval, for a total of 4 cycles. For high-throughput grinding, the TGrinder LH96 can be used in combination with a 96-well grinding tube adapter (TIANGEN, Cat. no. OSE-TH-A05); For large-volume samples, various stainless steel jars (OSE-TH-A08 to 12) are available. Set oscillation frequency at 60 Hz and grind for 2 min with steel beads.

**Note:** When using homogenizers from other manufacturers, optimize grinding parameters in accordance with the instrument's official protocol.

- 2.2 Add 800  $\mu$ l Buffer GPH (**confirm NKY tablets have been added beforehand**), 50  $\mu$ l Buffer ST and 20  $\mu$ l Proteinase K to the sample. Vortex thoroughly (TGyrate Master Vortex, Cat. no. OSE-VX-02 with accessory OSE-VX-M131 is recommended). Incubate at 60°C for 10 min (TGrade Dry Bath Incubator, OSE-DB series are recommended). Invert the tube several times during incubation for sufficient sample mixing.

**Note:**

- 1) After NKY tablets are added into Buffer GPH and vortexed for 2 min, a white insoluble suspension forms. Mix thoroughly before each use. Slight uneven distribution when aliquoting into centrifuge tubes will not affect the experimental results.
- 2) Buffer GPH, Buffer ST and Proteinase K can be premixed proportionally right before use. Do not store the mixed solution for extended periods.

2.3 Centrifuge at 12,000 rpm (~13,400×g) for 3 min.

**Note: Pipette only 600 µl supernatant for downstream experiments to avoid aspirating bottom impurities, discard residual liquid.**

### 3. Operating Steps for TGuide S16/S32 Pro/S96 Dex Nucleic Acid Extractor

3.1 Add 600 µl of the processed supernatant to wells in columns 1 and 7 of the 96-well deep plate. Add 5 µl of RNaseA (100 mg/ml) to columns 2 and 8.

**Note: To avoid affecting the activity of RNaseA, add RNaseA to columns 2 and 8 immediately before use. Do not pre-add; prepare fresh for each run.**

3.2 Place the 96-well deep plate onto the base platform inside TGuide S16/S32 Pro/S96 Dex instrument; insert tip combs into corresponding comb rack slots and fully lock the buckles in place.

3.3 Enter the program of the TGuide S16/ S32 Pro/S96 Dex, select the matching preset protocol file and click the Run button to start the experiment.

The detailed experimental procedures are shown in the table below:

Table 1: Hi-Plant DNA Extraction Program for TGuide S16 Automated Nucleic Acid Purification System

| Step | Hole site | Step name      | Mix time (min) | Mix speed | Dry time (min) | Volume (μl) | Temp. (°C) | Segments | Every time (sec) | Magnetization time(sec) | Cycle | Magnet speed (mm/s) |
|------|-----------|----------------|----------------|-----------|----------------|-------------|------------|----------|------------------|-------------------------|-------|---------------------|
| 1    | 6         | Transfer Beads | 0              | 8         | --             | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 2    | 2         | Retain Beads   | 1              | 8         | --             | 900         | --         | 0        | 0                | 0                       | 0     | 2.5                 |
| 3    | 1         | Bind           | 5              | 8         | --             | 900         | --         | 0        | 0                | 0                       | 0     | 2.5                 |
| 4    | 2         | Transfer Beads | 0              | 8         | --             | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 5    | 1         | Bind           | 5              | 8         | --             | 900         | --         | 5        | 10               | 0                       | 2     | 2.5                 |
| 6    | 2         | Wash 1         | 0.5            | 8         | --             | 100         | --         | 1        | 0                | 0                       | 1     | 2.5                 |
| 7    | 2         | Wash 2         | 6              | 8         | --             | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 8    | 3         | Wash 3         | 0.5            | 8         | --             | 100         | --         | 1        | 0                | 0                       | 1     | 2.5                 |
| 9    | 3         | Wash 4         | 2.5            | 8         | --             | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 10   | 4         | Wash 5         | 0.5            | 8         | --             | 100         | --         | 1        | 0                | 0                       | 1     | 2.5                 |
| 11   | 4         | Wash 6         | 2              | 8         | --             | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 12   | 6         | Wash 7         | 0.5            | 8         | --             | 100         | --         | 1        | 0                | 0                       | 1     | 2.5                 |
| 13   | 6         | Wash 8         | 2              | 8         | 6              | 900         | --         | 5        | 5                | 0                       | 2     | 2.5                 |
| 14   | 5         | Elution        | 6              | 8         | --             | 100         | 75         | 5        | 12               | 0                       | 2     | 2.5                 |
| 15   | 6         | Discard        | 0.5            | 8         | --             | 300         | --         | 1        | 0                | 0                       | 1     | 2.5                 |

Table 2: Hi-Plant DNA Extraction Program for TGuide S32 Pro Automated Nucleic Acid Purification System

| Step | Hole Site | Step Name      | Waiting time (min) | Mixing time (min) | Magnetic suction time (min) | Mixing speed | Volume (μl) | Temperature (°C) | Adsorption mode |
|------|-----------|----------------|--------------------|-------------------|-----------------------------|--------------|-------------|------------------|-----------------|
| 1    | 6         | Transfer Beads | 0                  | 0.5               | 1                           | Fast         | 910         | --               | Cycle           |
| 2    | 2         | Retain Beads   | 0                  | 0.5               | 0                           | Fast         | 900         | --               | ---             |
| 3    | 1         | Lysis          | 0                  | 5                 | 0                           | Fast         | 900         | --               | ---             |
| 4    | 2         | Transfer Beads | 0                  | 0                 | 1                           | Fast         | 900         | --               | Cycle           |
| 5    | 1         | Mix            | 5                  | 5                 | 1                           | Fast         | 900         | --               | Cycle           |
| 6    | 2         | Wash 1         | 0                  | 0.5               | 1                           | Fast         | 100         | --               | ---             |
| 7    | 2         | Wash 2         | 0                  | 6                 | 1                           | Fast         | 900         | --               | Cycle           |
| 8    | 3         | Wash 3         | 0                  | 0.5               | 1                           | Fast         | 100         | --               | ---             |
| 9    | 3         | Wash 4         | 0                  | 2.5               | 1                           | Fast         | 900         | --               | Cycle           |
| 10   | 4         | Wash 5         | 0                  | 0.5               | 1                           | Fast         | 100         | --               | ---             |
| 11   | 4         | Wash 6         | 0                  | 2                 | 1                           | Fast         | 900         | --               | Cycle           |
| 12   | 6         | Wash 7         | 0                  | 0.5               | 1                           | Fast         | 100         | --               | ---             |
| 13   | 6         | Wash 8         | 0                  | 2                 | 1                           | Fast         | 900         | --               | Cycle           |
| 14   | 5         | Elution        | 6                  | 6                 | 2                           | Fast         | 100         | 75               | Cycle           |
| 15   | 6         | Discard        | 0                  | 0.5               | 0                           | Fast         | 910         | --               | --              |

**Table 3: Hi-Plant DNA Extraction Program for TGuide S96 Dex Automated Nucleic Acid Purification System**

| Step | Hole Step | Step Name      | Waiting time before mixing (mm:ss) | Mixing speed | Mixing time (mm:ss) | Mixing mode | Liquid volume (μl) | Magnetic suction time (mm:ss) | Number of magnetic aspirations | Heating temperature (°C) | Pause after completion |
|------|-----------|----------------|------------------------------------|--------------|---------------------|-------------|--------------------|-------------------------------|--------------------------------|--------------------------|------------------------|
| 1    | 6         | Transfer Beads | 0                                  | Fast         | 0:30                | Normal Mode | 910                | 0:30                          | 2                              | -                        | -                      |
| 2    | 2         | Retain Beads   | 0                                  | Fast         | 0:30                | Normal Mode | 900                | 0:00                          | 0                              | -                        | -                      |
| 3    | 1         | Mix            | 0                                  | Fast         | 5:00                | Normal Mode | 900                | 0:00                          | 0                              | -                        | -                      |
| 4    | 2         | Transfer Beads | 0                                  | Fast         | 0:30                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 5    | 1         | Bind           | 0                                  | Fast         | 5:00                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 6    | 2         | Wash 1         | 0                                  | Fast         | 0:30                | Normal Mode | 100                | 0:30                          | 0                              | -                        | -                      |
| 7    | 2         | Wash 2         | 0                                  | Fast         | 6:00                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 8    | 3         | Wash 3         | 0                                  | Fast         | 0:30                | Normal Mode | 100                | 0:30                          | 0                              | -                        | -                      |
| 9    | 3         | Wash 4         | 0                                  | Fast         | 2:30                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 10   | 4         | Wash 5         | 0                                  | Fast         | 0:30                | Normal Mode | 100                | 0:30                          | 0                              | -                        | -                      |
| 11   | 4         | Wash 6         | 0                                  | Fast         | 2:00                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 12   | 6         | Wash 7         | 0                                  | Fast         | 0:30                | Normal Mode | 100                | 0:30                          | 0                              | -                        | -                      |
| 13   | 6         | Wash 8         | 0                                  | Fast         | 2:00                | Normal Mode | 900                | 0:30                          | 2                              | -                        | -                      |
| 14   | 5         | Elution        | 6:00                               | Fast         | 6:00                | Normal Mode | 100                | 0:30                          | 3                              | -                        | -                      |
| 15   | 6         | Discard        | 0                                  | Fast         | 0:30                | Normal Mode | 910                | 0                             | 0                              | -                        | -                      |

3.4 After completion of automated extraction program, aspirate purified DNA from columns 5 and 11 of the 96-well deep plate and store the DNA under appropriate conditions.

## Appendix

### Related Products

#### 1. Instrument and Accessories

| Product name                      | Packing Size   | Cat.No      |
|-----------------------------------|----------------|-------------|
| TGuide S16 Nucleic Acid Extractor | 1 set          | OSE-S16-AM  |
| TGuide Smart Magnetic Tip Comb    | 200 pieces/box | OSE-TGA-S03 |
| TGuide Single Sample Tank Bracket | 5 pieces/box   | OSE-TGA-S32 |

#### 2. TGuide Smart Reagents Kits

| Product name                          | Cartridge | Cat.No    |
|---------------------------------------|-----------|-----------|
| TGuide Smart Viral DNA/RNA Kit        | 48 preps  | GDP604-DE |
| TGuide Smart Universal DNA Kit        | 48 preps  | GDP605-DE |
| TGuide Smart Envir-DNA Kit            | 48 preps  | GDP613-DE |
| TGuide Smart Hi-Plant DNA Kit         | 48 preps  | GDP617-DE |
| TGuide Smart DNA Purification Kit     | 48 preps  | GDP642-DE |
| TGuide Smart Micro Cell Total RNA Kit | 48 preps  | GDP670-DE |
| TGuide Smart Universal RNA Kit        | 48 preps  | GDP671-DE |
| TGuide Smart Hi-Plant RNA Kit         | 48 preps  | GDP672-DE |
| TGuide Smart Processed Food DNA Kit   | 48 preps  | GDP680-DE |
| TGuide Smart Viral DNA/RNA Kit        | 96 preps  | GDP604-E  |
| TGuide Smart Universal DNA Kit        | 96 preps  | GDP605-E  |
| TGuide Smart Envir-DNA Kit            | 96 preps  | GDP613-E  |
| TGuide Smart Hi-Plant DNA Kit         | 96 preps  | GDP617-E  |
| TGuide Smart DNA Purification Kit     | 96 preps  | GDP642-E  |
| TGuide Smart Micro Cell Total RNA Kit | 96 preps  | GDP670-E  |
| TGuide Smart Universal RNA Kit        | 96 preps  | GDP671-E  |
| TGuide Smart Hi-Plant RNA Kit         | 96 preps  | GDP672-E  |
| TGuide Smart Processed Food DNA Kit   | 96 preps  | GDP680-E  |