



(DP305) Plant Genomic DNA Kit

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Experiment Preparation

1. Plant leaves; Mortar; Liquid nitrogen
2. 96-100% ethanol; Mercaptoethanol
3. Pipette and matched sterile tips (10 μ l, 200 μ l, 1 ml); 1.5 ml centrifuge tubes
4. Vortex oscillator; Dry bath/water bath; Centrifuge



Experiment Preparation-Kit Preparation 1

Please add 96-100% ethanol in Buffer PW and GD before use according to the volume indicated on the label of the bottle.



Experiment Preparation-Kit Preparation 1



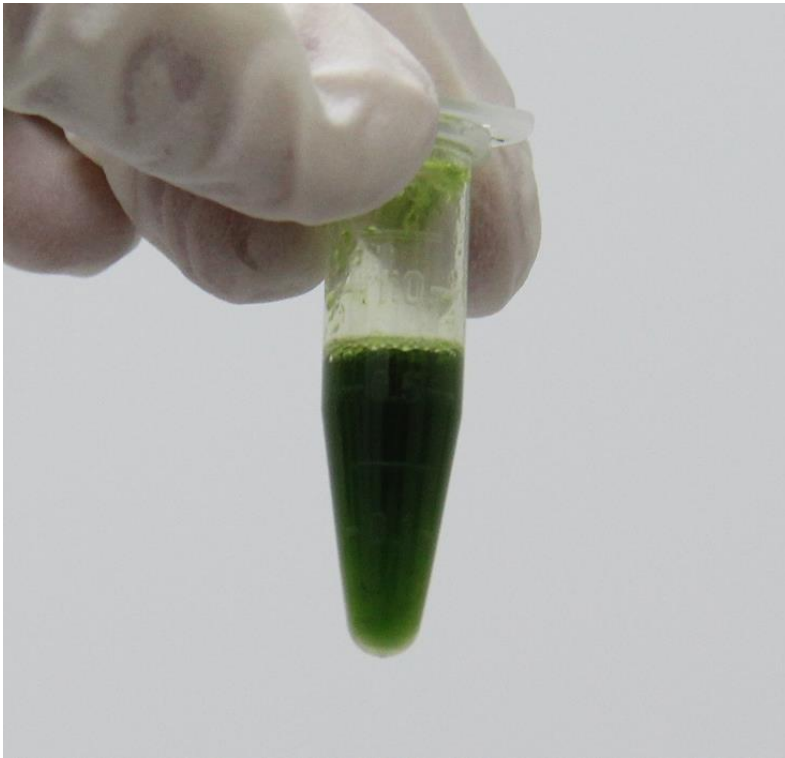
In preparation for the experiment, preheat Buffer GP1 in 65 °C. Add mercaptoethanol in preheating Buffer GP1 to make its final concentration to 0.1% before the experiment.

Step 1



Add liquid nitrogen to 100 mg fresh plant tissue or 30 mg dry weight tissue, and fully grind the tissues.

Step 2

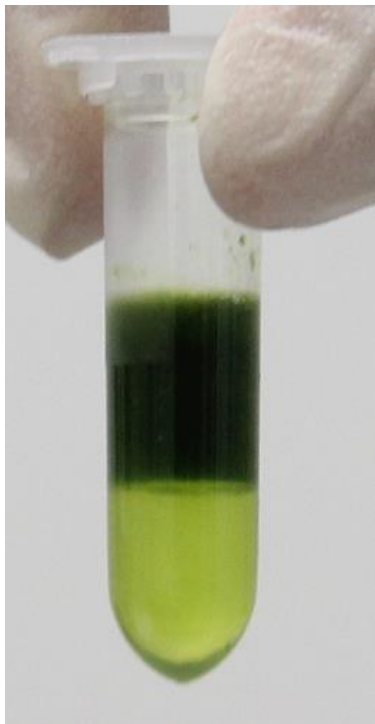


Quickly transfer the ground powder to the centrifugal tube containing 700 μ l preheating in 65 °C Buffer GP1.

Note: It is recommended to transfer with a metal spoon, which must be pre-cooled with liquid nitrogen before contacting the sample. Add mercaptoethanol in preheating Buffer GP1 to make its final concentration to 0.1% before the experiment.

Rapidly mix upside down, and then put the centrifuge tube in 65 °C dry bath/water bath for 20 min. In this process, mix the sample by turning the centrifuge tube upside down for several times.

Step 3



Add 700 μ l chloroform



Mix well



Centrifuge at 12,000 rpm
($\sim 13,400\times g$) for 5 min

Step 4



Carefully transfer the upper water phase from the previous step into a new centrifuge tube.

Add 700 μ l Buffer GP2 and mix well.

Step 5



Transfer the mixed liquid into Spin Column CB3.



Centrifuge at 12,000 rpm (~13,400×g) for 30 sec, and discard the waste liquid (The volume of the spin column is about 700 μl, so add liquid to centrifuge in different times).

Step 6



Add 500 μ l Buffer GD to Spin Column CB3



Centrifuge at 12,000 rpm ($\sim 13,400\times g$) for 30 sec, discard the waste liquid in the collection tube and place the Spin Column CB3 into the collection tube.

Step 7



Centrifuge at 12,000 rpm (~13,400×g) for 30 sec, discard waste liquid in the collection tube and place the Spin Column CB3 into the collection tube.

Add 600 µl Buffer PW to Spin Column CB3 (ensure 96-100% ethanol has been added before use).

Step 8 Repeat step 7.

Step 9



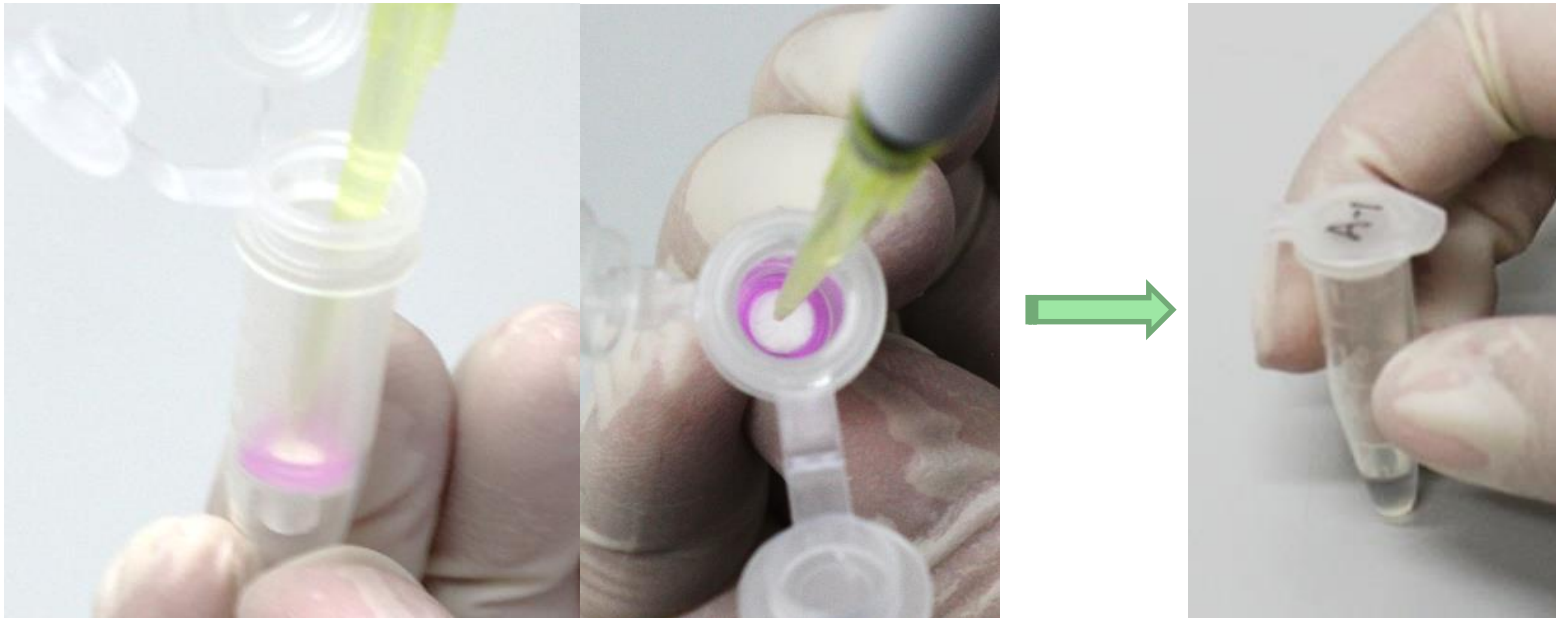
Centrifuge at 12,000 rpm ($\sim 13,400\times g$) for 2 min, and discard the waste liquid.



Place the Spin Column CB3 at room temperature for 2 minutes to completely dry Buffer PW in the membrane.

Note: Ethanol residues in Buffer PW can inhibit subsequent enzymatic reactions (restriction enzyme digestion, PCR, etc.) experiments.

Step 10



Transfer the Spin Column CB3 into a 1.5 ml centrifuge tube, and add 50-200 μ l Buffer TE to the middle of the adsorption membrane. Place at room temperature for 2 min and centrifuge at 12,000 rpm ($\sim 13,400\times g$) for 2 min to collect the solution into the centrifuge tube.