



# (DP348) TIANamp Blood DNA Kit

## -≤200 μl Anticoagulant whole blood

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# Experiment Preparation

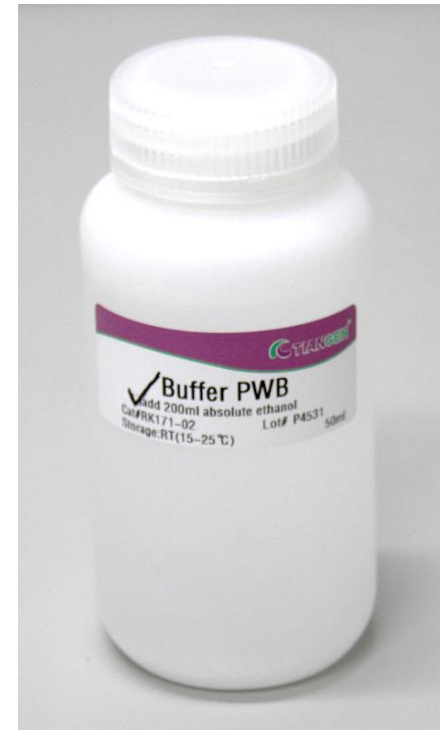
1. Anticoagulant blood ( $\leq 200 \mu\text{l}$ . Top up with Buffer GS to  $200 \mu\text{l}$ )
2. Pipette and matched sterile tips ( $200 \mu\text{l}$ ,  $1 \text{ ml}$ )
3. 96-100% ethanol
4. Vortex oscillator; Dry bath/water bath; Centrifuge



Note: This experiment takes human blood as an example. The whole blood of mammals can be extracted by this process. For anticoagulant blood of poultry, birds, amphibians or lower organisms, the red blood cells are nucleated cells, so the starting sample volume is  $5\text{-}20 \mu\text{l}$ , and Buffer GS should be added to top up to  $200 \mu\text{l}$ . For blood clot samples, the Liquefaction Columns CX1 (TIANGEN, RK165) (self-provided) can be selected to treat the blood clot.

# Experiment Preparation - Kit Preparation

Please add 96-100% ethanol in Buffer PWB before use according to the volume indicated on the label of the bottle.



# Step 1



When 200  $\mu$ l blood sample is extracted, carry out the next step directly. For blood sample less than 200  $\mu$ l, top up with Buffer GS to 200  $\mu$ l.

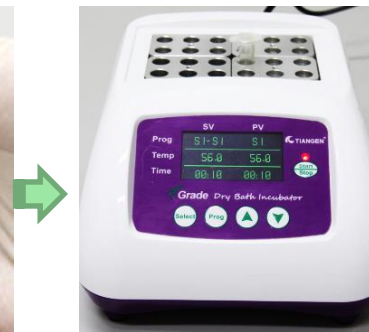
## Step 2



Add 200  $\mu$ l Buffer GB  
and 20  $\mu$ l Proteinase K.



Mix well upside  
down.



Incubate at 56°C  
for 10 min.



The solution should  
become clear at this  
step.

## Step 3



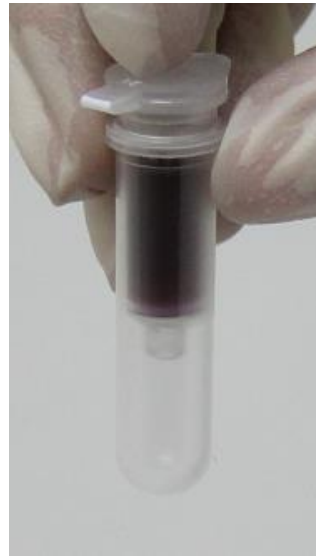
Incubate at room temperature for 2-5 min, then carefully add 350  $\mu$ l Buffer BD.

Mix well upside down, and there may be flocculent precipitate at this step.

## Step 4



Place Spin Column CG2 in a collection tube.



Transfer the solution and flocculent precipitate from the previous step to Spin Column CG2.



Centrifuge at 12,000 rpm (~13,400×g) for 30 sec. Discard the waste liquid in the collection tube and place the Spin Column CG2 back in the collection tube.

## Step 5



Add 500  $\mu$ l Buffer GDB to Spin Column CG2. Centrifuge at 12,000 rpm ( $\sim$ 13,400 $\times$ g) for 30 sec, discard the waste liquid in the collection tube and place the Spin Column CG2 into the collection tube.



## Step 6



Add 600  $\mu$ l Buffer PWB to Spin Column CG2 (ensure 96-100% ethanol has been added before use).

Centrifuge at 12,000 rpm ( $\sim 13,400\times g$ ) for 30 sec, discard waste liquid in the collection tube and place the Spin Column CG2 into the collection tube.

## Step 7 Repeat step 6.

## Step 8

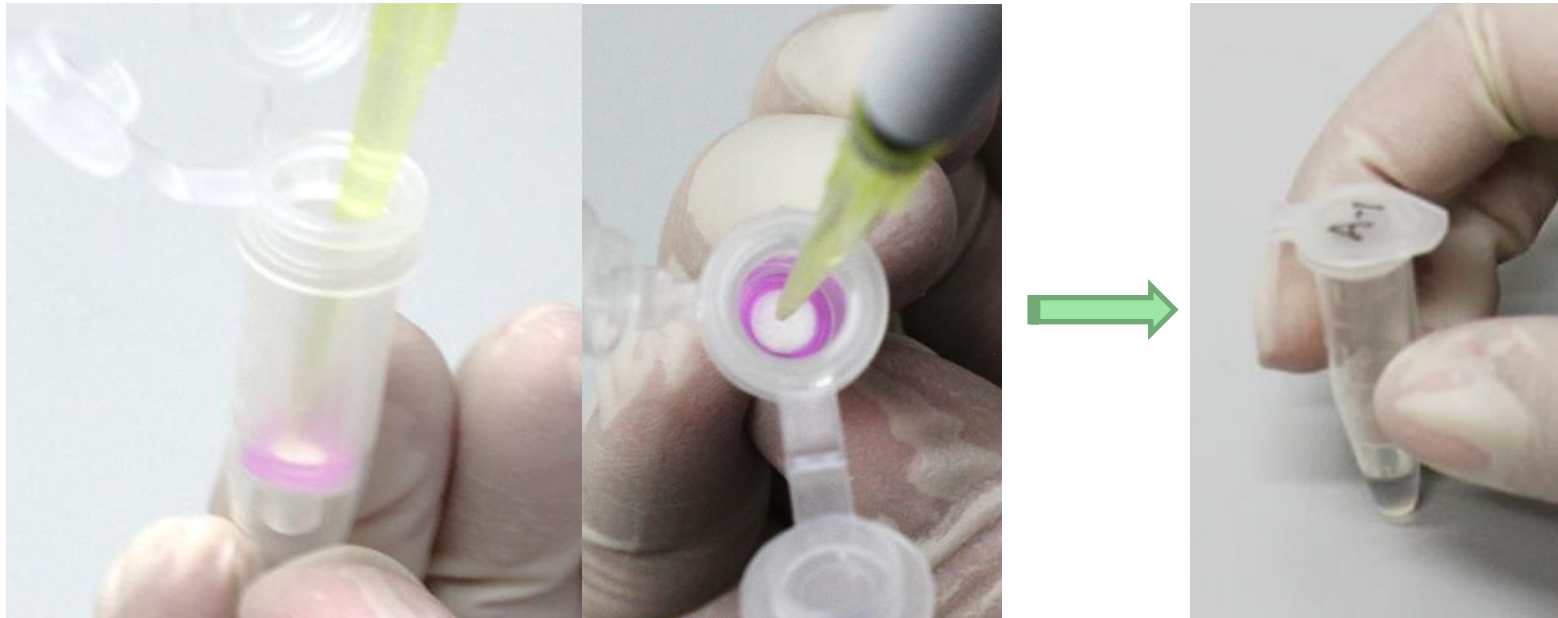


Centrifuge at 12,000 rpm ( $\sim 13,400\times g$ ) for 2 min, and discard the waste liquid.

Place the Spin Column CG2 at room temperature for 2 minutes to completely dry Buffer PWB in the membrane.

**Note:** Ethanol residues in Buffer PWB can inhibit subsequent enzymatic reactions (restriction enzyme digestion, PCR, etc.) experiments.

## Step 9



Transfer the Spin Column CG2 into a 1.5 ml centrifuge tube, and add 50-200  $\mu$ l Buffer TB to the middle of the adsorption membrane. Place at room temperature for 2 min and centrifuge at 12,000 rpm ( $\sim 13,400\times g$ ) for 2 min to collect the solution into the centrifuge tube.